



## Ciliates – Protists with complex morphologies and ambiguous early fossil record



Micah Dunthorn <sup>a,\*</sup>, Jere H. Lipps <sup>b,c</sup>, John R. Dolan <sup>d</sup>, Marie Abboud-Abi Saab <sup>e</sup>, Erna Aesch <sup>f</sup>, Charles Bachy <sup>g</sup>, María Sonia Barría de Cao <sup>h</sup>, Helmut Berger <sup>i</sup>, William A. Bourland <sup>j</sup>, Joong Ki Choi <sup>k</sup>, John Clamp <sup>l</sup>, Mary Doherty <sup>m</sup>, Feng Gao <sup>n</sup>, Eleni Gentekaki <sup>o</sup>, Jun Gong <sup>p</sup>, Xiaozhong Hu <sup>q</sup>, Jie Huang <sup>r</sup>, Takashi Kamiyama <sup>s</sup>, Matthew D. Johnson <sup>t</sup>, Barbara Kammerlander <sup>u,v</sup>, Sun Young Kim <sup>w</sup>, Young-Ok Kim <sup>x</sup>, Antonietta la Terza <sup>y</sup>, Michèle Laval-Peuto <sup>z</sup>, Diana Lipscomb <sup>aa</sup>, Christopher S. Lobban <sup>ab</sup>, Hongan Long <sup>ac</sup>, Pierangelo Luporini <sup>y</sup>, Denis H. Lynn <sup>ad</sup>, Miroslav Macek <sup>ae</sup>, Robert I. Mansergh <sup>af</sup>, Mercedes Martín-Cereceda <sup>ag</sup>, George G. McManus <sup>ah</sup>, David J.S. Montagnes <sup>ai</sup>, Geoffrey O. Ong'ondo <sup>aj</sup>, David J. Patterson <sup>ak</sup>, Blanca Pérez-Uz <sup>ag</sup>, Pablo Quintela-Alonso <sup>al,ag</sup>, Lúcia S.L. Safi <sup>am</sup>, Luciana F. Santoferrara <sup>ah</sup>, Bettina Sonntag <sup>u</sup>, Weibo Song <sup>n</sup>, Thorsten Stoeck <sup>a</sup>, Diane K. Stoecker <sup>an</sup>, Michaela C. Strüder-Kypke <sup>ao</sup>, Isabelle Trautmann <sup>a</sup>, Laura R.P. Utz <sup>ap</sup>, Adriana Vallesi <sup>y</sup>, Peter Vd'ačný <sup>aq</sup>, Alan Warren <sup>ar</sup>, Thomas Weisse <sup>u,as</sup>, Stephen A. Wickham <sup>at</sup>, Zhenzhen Yi <sup>au</sup>, Wuchang Zhang <sup>av</sup>, Zifeng Zhan <sup>av</sup>, Rebecca Zufall <sup>aw</sup>, Sabine Agatha <sup>at,\*</sup>

<sup>a</sup> Department of Ecology, University of Kaiserslautern, Kaiserslautern, Germany

<sup>b</sup> University of California Museum of Paleontology, Berkeley, CA, USA

<sup>c</sup> John D. Cooper Archaeological and Paleontological Center, Santa Ana, CA, USA

<sup>d</sup> Laboratoire d'Océanographie de Villefranche-sur-Mer, CNRS Université Paris VI, Villefranche-sur-Mer, France

<sup>e</sup> CNRS/National Center for Marine Sciences, Batroun, Lebanon

<sup>f</sup> Oberösterreichisches Landesmuseum Biologiezentrum, Linz, Austria

<sup>g</sup> Monterey Bay Aquarium Research Institute, Moss Landing, CA, USA

<sup>h</sup> Instituto Argentino de Oceanografía, Bahía Blanca, Argentina

<sup>i</sup> Consulting Engineering Office for Ecology, Salzburg, Austria

<sup>j</sup> Department of Biological Sciences, Boise State University, Boise, ID, USA

<sup>k</sup> Department of Oceanography, Inha University, Incheon, South Korea

<sup>l</sup> North Carolina Central University, Durham, NC, USA

<sup>m</sup> Department of Biology, Rhodes College, Memphis, TN, USA

<sup>n</sup> Laboratory of Protozoology, Ocean University of China, Qingdao, China

<sup>o</sup> Department of Biology, Chulalongkorn University, Bangkok, Thailand

<sup>p</sup> Yantai Institute of Coastal Zone Research, Chinese Academy of Sciences, Yantai, China

<sup>q</sup> Institute of Evolution and Marine Biodiversity, Ocean University of China, Qingdao, China

<sup>r</sup> Key Laboratory of Aquatic Biodiversity and Conservation of Chinese Academy of Sciences, Institute of Hydrobiology, Chinese Academy of Sciences, Wuhan, China

<sup>s</sup> Tohoku National Fisheries Research Institute, Fisheries Research Agency, Japan

<sup>t</sup> Woods Hole Oceanographic Institution, Woods Hole, MA, USA

<sup>u</sup> Research Institute for Limnology, University of Innsbruck, Mondsee, Austria

<sup>v</sup> Institute of Ecology, University of Innsbruck, Innsbruck, Austria

<sup>w</sup> Resources Research Division, The National Institute of Marine Biological Resources Planning Bureau, Ministry of Oceans and Fisheries, South Korea

<sup>x</sup> Korean Institute of Ocean Science and Technology, South Korea

<sup>y</sup> School of Bioscience and Veterinary Medicine, University of Camerino, Camerino, Italy

<sup>z</sup> Chemin des Campons 144, La Colle-sur-Loup, France

<sup>aa</sup> Department of Biological Sciences, George Washington University, Washington DC, USA

<sup>ab</sup> Division of Natural Sciences, University of Guam, Mangilao, GU, USA

<sup>ac</sup> Department of Biology, Indiana University, Bloomington, IN, USA

<sup>ad</sup> Department of Zoology, University of British Columbia, Vancouver, BC, Canada

<sup>ae</sup> Facultad de Estudios Superiores Iztacala, Universidad Nacional Autónoma de México, Tlalnepantla, México

<sup>af</sup> Institute of Marine Sciences, University of Portsmouth, United Kingdom

<sup>ag</sup> Departamento de Microbiología III, Facultad de Ciencias Biológicas, Universidad Complutense de Madrid, Spain

<sup>ah</sup> Department of Marine Sciences, University of Connecticut, Groton, CT, USA

<sup>ai</sup> Institute of Integrative Biology, University of Liverpool, Liverpool, United Kingdom

<sup>aj</sup> Department of Biological Sciences, Egerton University, Egerton, Kenya

<sup>ak</sup> School of Biological Sciences, University of Sydney, Australia

\* Corresponding authors.

E-mail addresses: [dunthorn@rhrk.uni-kl.de](mailto:dunthorn@rhrk.uni-kl.de) (M. Dunthorn), [sabine.agatha@sbg.ac.at](mailto:sabine.agatha@sbg.ac.at) (S. Agatha).

<sup>a1</sup> Department of General Ecology, Cologne Biocenter, University of Cologne, Cologne, Germany

<sup>am</sup> Virginia Institute of Marine Science, The College of William & Mary, VA, USA

<sup>an</sup> University of Maryland Center for Environmental Science, Cambridge, MD, USA

<sup>ao</sup> Department of Molecular and Cell Biology, University of Guelph, Guelph, ON, Canada

<sup>ap</sup> Faculty of Biosciences, Pontifícia Universidade Católica do Rio Grande do Sul (PUCRS), Rio Grande do Sul, Brazil

<sup>aq</sup> Department of Zoology, Comenius University, Bratislava, Slovak Republic

<sup>ar</sup> Department of Life Sciences, Natural History Museum, London, United Kingdom

<sup>as</sup> Editor of the European Journal of Protistology, Austria

<sup>at</sup> Department of Ecology and Evolution, University of Salzburg, Austria

<sup>au</sup> South China Normal University, Guangzhou, China

<sup>av</sup> Institute of Oceanology, Chinese Academy of Sciences, Qingdao, China

<sup>aw</sup> Department of Biology and Biochemistry, University of Houston, Houston, TX, USA

## ARTICLE INFO

### Article history:

Received 3 October 2014

Received in revised form 9 April 2015

Accepted 18 May 2015

Available online 5 June 2015

### Keywords:

Acritarch

Ciliophora

Taxonomy

Tintinnids

## ABSTRACT

Since ciliates rarely possess structures that easily fossilize, we are limited in our ability to use paleontological studies to reconstruct the early evolution of this large and ecologically important clade of protists. Tintinnids, a group of loricate (house-forming) planktonic ciliates, are the only group that has a significant fossil record. Putative tintinnid fossils from rocks older than Jurassic, however, possess few to no characters that can be found in extant ciliates; these fossils are best described as ‘*incertae sedis* eukaryotes’. Here, we review the Devonian fossil *Nassacysta reticulata* and propose that it is likewise another *incertae sedis* eukaryote due to the lack of any unambiguous ciliate characters. Future tintinnid fossil descriptions would be most helpful if: (i) neutral terminology is used in the descriptions but ciliate-specific terminology in the interpretations; (ii) the current ciliate classification is used, although fossil data may expand or modify classifications based on modern forms; (iii) close collaboration with specialists studying extant ciliates is done; and (iv) editors include an expert of extant ciliates in the review process.

© 2015 Elsevier B.V. All rights reserved.

## Contents

1. Introduction . . . . .	2
2. Non-ciliate proterozoic and paleozoic microfossils . . . . .	2
3. Moving forward with future putative ciliate fossils . . . . .	4
Acknowledgments . . . . .	5
References. . . . .	5

## 1. Introduction

Ciliates (Alveolata, Ciliophora) are morphologically extremely diverse in comparison with most other protists (Dunthorn and Katz, 2008; Lynn, 2008). Species and higher taxa are primarily distinguished by variations in cell shape and size, the numbers and patterns of oral and somatic cilia, and the ultrastructure of the fibers associated with the somatic basal bodies underlying the cilia. These minute characters are observed in live and silver-stained specimens, as well as with electron microscopy (Foissner, 2014; Lynn, 2008). Ciliates generally lack hard structures. The prospect for the fossilization of most of the cellular characters that are needed for unambiguous identification and description is therefore extremely low, although overall soft-cell shapes can be fossilized under a few conditions such as in amber (Ascaso et al., 2005; Martín-González et al., 2008; Poinar et al., 1993; Schmidt et al., 2006; Schönborn et al., 1999).

Tintinnid ciliates (Spirotricha, Choreotrichida, Tintinnina) have external loricae (Fig. 1), which are more easily fossilized and have a long, but patchy, history in the rock record (Colom, 1948; Lipps et al., 2013; Tappan, 1993; Tappan and Loeblich, 1968). These fossils reliably identified as tintinnids—at least in terms of having the same lorica shape and composition as extant species—first appeared in the Jurassic and range to the Recent (Lipps et al., 2013; Remane, 1985; Rüst, 1885). Some other ciliates (e.g., colpodeans, peritrichs, and trachelophyllids) have loricae, cysts or scales that can potentially be fossilized (Lynn, 2008), although many other protist groups have similar structures as well (e.g., testate amoebae and foraminifera; Hausmann et al., 2003). The hydrocarbon gammacerane was posited to be a molecular fossil of

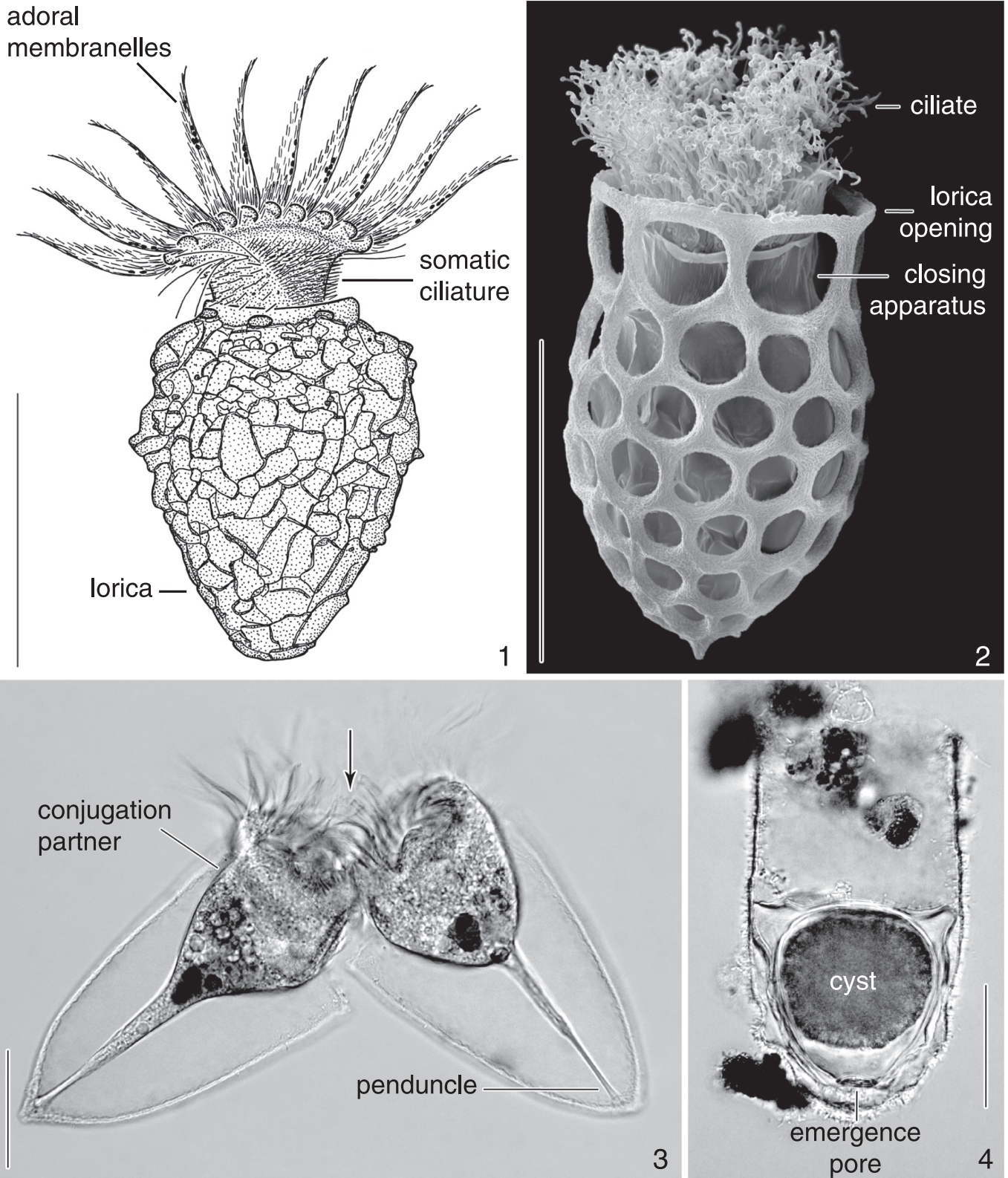
ciliates as its precursor, tetrahymanol, was thought to be found only in *Tetrahymana* (Summons and Walter, 1990); but tetrahymanol was recently discovered in a variety of organisms (Takishita et al., 2012).

As a community of ciliate researchers who largely study extant ciliates, we look forward to any new and accurately identified ciliate fossil, especially if it originates from pre-Jurassic rocks. Because most genes in ciliates suffer from extensive paralogy and excessive nucleotide substitutions, we are limited in our ability to use molecular data from extant species to infer early evolution and deep relationships (Dunthorn et al., 2014; Lynn, 2008). Publication of new ciliate fossils therefore can have profound effects on how we interpret the history of this large and ecologically important clade of protists.

## 2. Non-ciliate proterozoic and paleozoic microfossils

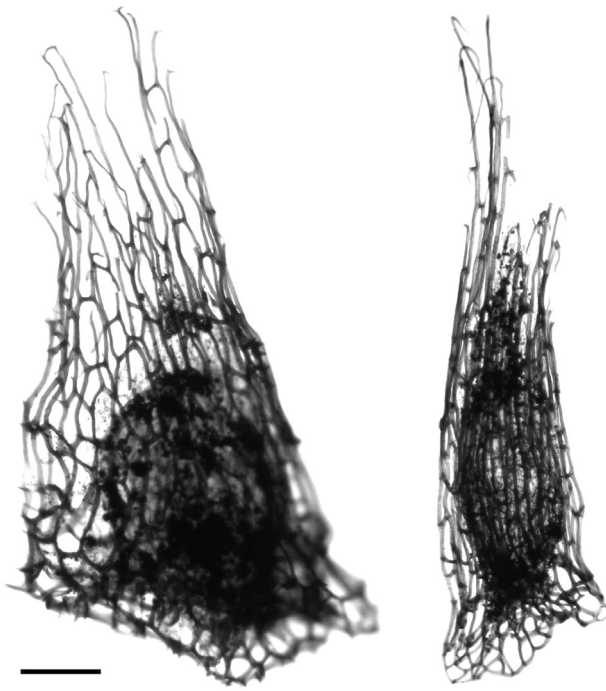
Molecular clock analyses place the origin of ciliates sometime in the Proterozoic (Berney and Pawlowski, 2006; Eme et al., 2014). Several fossils from Proterozoic rocks have recently been described as ciliates (Bosak et al., 2011; Li et al., 2007, 2009; Li and Zhang, 2006). As discussed elsewhere (Dunthorn et al., 2010; Lipps et al., 2013), these fossils have characteristics that were immediately recognized by researchers working on extant ciliates, as well as a ciliate fossil expert, as being non-ciliate in origin. These purported Proterozoic ciliate fossils are best described as *incertae sedis* eukaryotes or perhaps even inorganic in origin in few cases. They are not ciliates.

A more recent example of a problematic description is that from a new set of fossils from Middle Devonian (Givetian) sedimentary rock in the Ghadamis Basin (ca. 383–389 MYA) of Libya that has been named



**Figs. 1–4.** Morphology of extant tintinnid ciliates. 1: *Stenosemella pacifica*, a typical tintinnid ciliate (modified from Agatha and Tsai, 2008). Extended live cell projecting its anterior end with the membranelles used for locomotion and filter feeding out of the lorica. The somatic (body) cilia are arranged in specialized fields and rows. The lorica is composed of a small hyaline collar with minute windows and an agglutinated bowl. 2: *Dictyocysta mitra*, one of the few ciliates with a lorica sac and a reticulate lorica wall (modified from Agatha, 2010). The lateral view shows the extended ciliate within the membranous lorica sac that lines the lorica. The lorica sac merges anteriorly into a closing apparatus that forms typical triangular folds in closed condition. 3: Conjugating specimens of the genus *Schmidingerella* (original). The arrow marks the cytoplasmic bridge through which the division products of the micronucleus (migratory nuclei) are exchanged. 4: Resting cyst in the genus *Schmidingerella* (original). Scale bars = 40  $\mu$ m.





**Fig. 5.** Two fossils of *Nassacysta reticulata* identified by Steemans et al. (2014) as tintinnid-like ciliates. Since this fossil (holotype left) displays numerous characters dissimilar of tintinnids and ciliates in general, it is best classified as *incertae sedis* eukaryote. Scale bar = 100  $\mu\text{m}$  (incorrectly labeled as 10  $\mu\text{m}$  in the original publication). Figures modified from Steemans et al. (2014) and copyrighted by Elsevier, used with permission.

*Nassacysta reticulata*, and interpreted as being a ciliate (Fig. 5; Steemans et al., 2014). These fossils are organic-walled and 564–1617  $\mu\text{m}$  in length. They are composed of two structures: “an outer sheath looking like a fish trap open at its narrowest tip; and an inner rounded body enclosed in a thin membrane” (Steemans et al., 2014). The sheath itself is composed of threads about 10  $\mu\text{m}$  wide that form a reticulate structure with polygonal to subcircular meshes. Some of the threads have flat, distally enlarged projections. A membranous structure is also described, but it only covers the threads. The inner round bodies are interpreted to be resting cysts (Steemans et al., 2014).

Steemans et al. (2014) interpreted *Nassacysta* to be tintinnid-like based on the presence of its vase-shaped lorica and the occurrence of an internal resting cyst. *Nassacysta* was therefore placed within the ciliates under “Tintinnidia? Kofoid and Campbell, 1929”, although this name has not been published as valid; they may have either meant Tintinnida Kofoid and Campbell, 1929 or Tintinnina Kofoid and Campbell, 1929.

Several problems exist with the interpretation of *Nassacysta* as being tintinnid-like. First, the fossils are larger than the vast majority of extant tintinnids, 564–1617  $\mu\text{m}$  vs. rarely more than 300  $\mu\text{m}$  in length (Dolan, 2010). The problem of being too large was sufficient for Steemans et al. (2014) to reject an affinity with dinoflagellate cysts, chitinozoa (an extinct group of chitinous, flask-shaped microfossils that may be related to metazoa), thecamoebae (testate amoebae), miospores (spores or pollen grains less than 200  $\mu\text{m}$  across) like *Retispora lepidophyta*, gymnosperm pollen, and acritarchs, but not for tintinnids.

Second, Steemans et al. (2014) described the *Nassacysta* fossils as being vase-shaped, with a broad base and narrowing at the tip with apparent breaks in the outer net-like structure. While almost all tintinnid loricae have a vase-like shape, importantly they have a narrower base and a broader apical aperture with a distinct opening rim, where the cells can easily move in and out (Agatha et al., 2013)—exactly opposite of *Nassacysta*. Only two extant tintinnids (*Tintinnidium mucicola* and *Tintinnopsis amphora*) have a somewhat narrowed apical portion, but neither has a reticulate lorica and both have typical lorica apertures.

Steemans et al. (2014) interpreted the typical lorica opening rim as weak and thus torn off sometime during the fossilization in all specimens found, resulting in these distinct breaks. The supposition that *Nassacysta* are tintinnid-like necessitates an unjustified further assumption of apical apertures with distinct rims that were never observed. A more parsimonious explanation for the missing tintinnid-like lorica opening rims in every *Nassacysta* fossil found is that they never existed.

Third, reticulate loricae occur only in about seven tintinnid species belonging to the genus *Dictyocysta* (Fig. 2), whose lorica apertures are, like those of the other one thousand known tintinnid species, wider than the cell’s apical oral region and are surrounded by a distinct rim. The outer sheath of *Nassacysta* differs from the loricae in *Dictyocysta* in several respects: the size and structure of the threads, 10  $\mu\text{m}$  wide and with a distinct middle lamella as shown in Figs. 9 and 11 in Steemans et al. (2014) vs. less than 5  $\mu\text{m}$  wide and composed of minute tubules; the shape of the windows, oblong polygonal vs. circular to elliptical; and the presence of distally flattened and broadened projections (“sticks”; present vs. absent) (Agatha, 2010; Laval-Peuto, 1994; Steemans et al., 2014).

Fourth, Steemans et al.’s (2014) interpretation of the enclosed globular structures in *Nassacysta* necessitates a simultaneous cyst formation in all cells right before commencement of the fossilization process. This interpretation fails to explain the absence of empty loricae, which are typically found in Recent sediments and sediment traps in much greater abundances than loricae enclosing resting cysts (Boltovskoy et al., 1996; González et al., 2004; Ling, 1992; Suzuki and Taniguchi, 1995). Accordingly, there is no evidence that they were active, swimming cells prior to encystment as suggested by Steemans et al. (2014). A more parsimonious explanation for these structures is that they perhaps represent wind-blown seeds of extinct plants or extant contaminants.

Fifth, Steemans et al. (2014) interpreted some of the laterally fused *Nassacysta* as having sex by stating: “when modern tintinnids are agglutinated, they are associated by joining their oral pole one to the other, while our specimens are laterally associated. However, the tintinnid life cycle is not well known (Tappan, 1993). Lateral fusion has been observed among other ciliate-like dinoflagellates during sexual reproduction...”. This statement about tintinnid sex is problematic for several reasons. Tintinnids typically fuse only in their anterior cell portions (close underneath their apical cell ends where their mouths are) to exchange haploid products of their micronuclei (called conjugation); this exchange is how ciliates have sex (Dunthorn and Katz, 2010; Lynn, 2008). Although the life cycles of most tintinnids have not been examined in detail, a cytoplasmic bridge must be formed between mating cells to allow for genetic exchange (Fig. 3); such exchange by soft cell structures cannot occur through the hard wall of the loricae. Additionally, ciliates and dinoflagellates are placed together in the Alveolata, but we are unaware of organisms that may be described as a “ciliate-like dinoflagellate”.

These obvious differences and the lack of unambiguous ciliate characters, which were already partially recognized by Steemans et al. (2014), immediately show the non-ciliate nature of these fossils to those who work on extant ciliates. These differences are more than enough to exclude placing *Nassacysta* among the tintinnids specifically and in the ciliates generally; these fossils are best considered an *incertae sedis* eukaryote.

### 3. Moving forward with future putative ciliate fossils

*Nassacysta* from the Ghadamis Basin (Steemans et al., 2014) now joins other purported ciliate fossils from the Doushantuo Formation (Li et al., 2007), the Huangmailing Formation (Li and Zhang, 2006; Li et al., 2009), and the Tsagaan Oloom Formation (Bosak et al., 2011), that are also immediately recognizable as being not ciliates by researchers working on extant species because of conspicuous morphological differences or the lack of any unambiguous morphological characters that would unite them with ciliates. We cannot identify them any further than as *incertae sedis* eukaryotes.

Although there may be impetus to place a new fossil into a taxon of extant organisms such as ciliates or other protists, especially if it is useful for stratigraphy, sometimes all that can be said is that it is a microfossil of uncertain eukaryotic affinity; e.g., as Cohen and Knoll (2012) did with an extensive collection of mid-Neoproterozoic scale microfossils. Forcing fossils into taxa without definite criteria to do so impedes prompt understanding of their evolutionary relationships and should be avoided. If a fossil is to be described as an extinct member of the ciliates, we—the active community of taxonomists, phylogeneticists, and ecologists working on extant ciliates—recommend the following steps.

We ask that care be taken in using the current ciliate-specific terminology (e.g., Lynn, 2008) when interpreting the morphological structures in the fossils, which will allow a firm comparison with extant taxa. We emphasize the application of these terms in the interpretation of the fossil structures, not in the description of the fossils that may better be done with plain language. Specifically, the loricae of extant tintinnids are easily recognizable by a combination of the following characters (Agatha, 2010; Agatha et al., 2013; Agatha and Simon, 2012). The lorica is a single subspherical, obconical, obovoidal, cylindrical, bell-shaped, or flask-shaped chamber; the posterior portion of the bowl might be tapered, and a flaring or cylindrical apical collar might be set off against the bowl. The apical opening of the lorica is wide enough to allow an unhampered extension and contraction of the wide apical oral region of the ciliate cells. Its rim is continuous, but might be somewhat irregular or with teeth or gutters. The loricae are derived from cell secretions to which foreign material may attach, resulting in entirely agglutinated loricae, loricae composed of a hyaline collar and agglutinated bowls, or entirely hyaline loricae. Hyaline loricae or lorica portions might have rounded windows, surface ridges, or fins. The resting cysts are ellipsoidal or flask-shaped and have a distinct wall and occasionally an emergence pore (Fig. 4). Knoll (2014) noted that systematic interpretation of fossils in ancient assemblages could include or even consist entirely of an extinct stem-group (= basal grade); therefore, some of the tintinnid lorica characters listed here may represent derived characters not occurring in fossils of the ancestors.

We ask that authors describing fossils as ciliates should use the current ciliate classification (e.g., Adl et al., 2012; Lynn, 2008) and carefully consider work reporting related evidence from extant species. This may be done by collaborating with someone working with extant species, and it would greatly facilitate a more accurate use of the terminology in the interpretations and taxonomic assignments. Such constructive cross-field collaborations occurred in Porter et al. (2003) with Neoproterozoic testate amoebae fossils and in Schönborn et al. (1999) with Triassic protists in amber.

Additionally, we ask authors and editors who are handling putative ciliate fossil submissions might include an expert of extant ciliates in the review process. In addition to comparing a new fossil with previously described ones, which paleontologists are in the best position to do especially if there have been taphonomic and diagenetic changes, comparisons with extant species are also essential. These steps could at least assist in the interpretations of putative ciliates found in the rock record.

## Acknowledgments

We thank Stefan Bengtson, Andrew H. Knoll, Jan Pawlowski, and an anonymous reviewer for constructive comments. Funding for this paper came from the Deutsche Forschungsgemeinschaft (grant # DU1319/1-1) to MD and the National Science Foundation (DEB grant # 1136580) to JC.

## References

Adl, S.M., Simpson, A.G., Lane, C.E., Lukeš, J., Bass, D., Bowser, S.S., Brown, M.W., Burki, F., Dunthorn, M., Hampl, V., Heiss, A., Hoppenrath, M., Lara, E., Le Gall, L., Lynn, D.H., McManus, H., Mitchell, E.A.D., Mozley-Stanridge, S.E., Parfrey, L.W., Pawlowski, J., Rueckert, S., Shadwick, L., Schoch, C., Smirnov, A., Spiegel, F.W., 2012. The revised classification of eukaryotes. *J. Eukaryot. Microbiol.* 59, 429–493.

Agatha, S., 2010. A light and scanning electron microscopic study of the closing apparatus in tintinnid ciliates (Ciliophora, Spirotricha, Tintinnina): a forgotten synapomorphy. *J. Eukaryot. Microbiol.* 57, 297–307.

Agatha, S., Simon, P., 2012. On the nature of tintinnid loricae (Ciliophora: Spirotricha: Tintinnina): a histochemical, enzymatic, EDX, and high-resolution TEM study. *Acta Protozool.* 51, 1–19.

Agatha, S., Tsai, S.-F., 2008. Redescription of the tintinnid *Stenosemella pacifica* Kofoid and Campbell, 1929 (Ciliophora, Spirotricha) based on live observation, protargol impregnation, and scanning electron microscopy. *J. Eukaryot. Microbiol.* 55, 75–85.

Agatha, S., Laval-Peuto, M., Simon, P., 2013. The Tintinnid Lorica. In: Dolan, J.R., Montagnes, D.J.S., Agatha, S., Coats, D.W., Stoecker, D.K. (Eds.), *The biology and ecology of tintinnid ciliates: models for marine plankton*. Wiley-Blackwell, West Sussex, pp. 17–41.

Ascaso, C., Wierzbos, J., Speranza, M., Gutiérrez, J.C., González, A.M., de los Ríos, A., Alonso, J., 2005. Fossil protists and fungi in amber and rock substrates. *Micropaleontology* 51, 59–72.

Berney, C., Pawlowski, J., 2006. A molecular time-scale for eukaryote evolution recalibrated with the continuous microfossil record. *Proc. R. Soc. Lond. B* 273, 1867–1872.

Boltovskoy, D., Uliana, E., Wefer, G., 1996. Seasonal variation in the flux of microplankton and radiolarian assemblage compositions in the northeastern tropical Atlantic at 2.195 m. *Limnol. Oceanogr.* 41, 615–635.

Bosak, T., Macdonald, F., Lahr, D., Matys, E., 2011. Putative Cryogenian ciliates from Mongolia. *Geology* 39, 1123–1126.

Cohen, P.A., Knoll, A.H., 2012. Neoproterozoic scale microfossils from the Fifteenmile Group, Yukon Territory. *J. Paleontol.* 86, 775–800.

Colom, G., 1948. Fossil tintinnids: loricated infusoria of the order of the Oligotricha. *J. Paleontol.* 22, 233–263.

Dolan, J.R., 2010. Morphology and ecology in tintinnid ciliates of the marine plankton: correlates of lorica dimensions. *Acta Protozool.* 49, 235–244.

Dunthorn, M., Katz, L.A., 2008. Richness of morphological hypotheses in ciliate systematics allows for detailed assessment of homology and comparisons with gene trees. *Denisia* 23, 389–394.

Dunthorn, M., Katz, L.A., 2010. Secretive ciliates and putative asexuality in microbial eukaryotes. *Trends Microbiol.* 18, 183–188.

Dunthorn, M., Lipps, J.H., Stoeck, T., 2010. Reassessment of the putative ciliate fossils *Eotintinnopsis*, *Wujiangella*, and *Yonyangella* from the Neoproterozoic Doushantuo Formation in China. *Acta Protozool.* 49, 139–144.

Dunthorn, M., Otto, J., Berger, S.A., Stamatakis, A., Mahé, F., Romac, S., de Vargas, C., Audic, S., BioMark Consortium, Stock, A., Kauff, F., Stoeck, T., 2014. Placing environmental next-generation sequencing amplicons from microbial eukaryotes into a phylogenetic context. *Mol. Biol. Evol.* 31, 993–1009.

Eme, L., Sharpe, S.C., Brown, M.W., Roger, A.J., 2014. On the age of eukaryotes: evaluating evidence from fossils and molecular clocks. *Cold Spring Harb. Perspect. Biol.* 6, a016139.

Foissner, 2014. An update of 'basic light and scanning electron microscopic methods for taxonomic studies of ciliated protozoa'. *Int. J. Syst. Evol. Microbiol.* 64, 271–292.

González, H.E., Hebbeln, D., Iriarte, J.L., Marchant, M., 2004. Downward fluxes of faecal material and microplankton at 2300 m depth in the oceanic area off Coquimbo (30° S), Chile, during 1993–1995. *Deep-Sea Res. II* 51, 2457–2474.

Hausmann, K., Hülsmann, N., Radek, R., 2003. *Protistology*. 3rd ed. E. Schweizerbart'sche Verlagsbuchhandlung, Berlin.

Knoll, A.H., 2014. Paleobiological perspectives on early eukaryotic evolution. *Cold Spring Harb. Perspect. Biol.* 6, a016121.

Laval-Peuto, M., 1994. Classe des Oligotricha Bütschli, 1887. *Ordre des Tintinnida Kofoid et Campbell, 1929. Traité de Zoologie. Anatomie, systématique, biologie. II. Infusaires ciliés 2. Systématique*. Masson, Paris, pp. 181–219.

Li, Y.-X., Zhang, S.-X., 2006. New material of microfossils from Middle Proterozoic in Hubei Province. *Acta Palaeontol. Sin.* 45, 102–107.

Li, C.-W., Chen, J.-Y., Lipps, J.H., Gao, F., Chi, H.-M., Wu, H.-J., 2007. Ciliated protozoans from the Precambrian Doushantuo Formation, Wengsen, South China. *Geol. Soc. Spec. Pub.* 286, 151–156.

Li, Y.-X., Zhang, S.-X., Zhang, J., 2009. Mesoproterozoic Calymnian tintinnids from central China. *Open Paleontol. J.* 2, 10–13.

Ling, H.Y., 1992. *Tintinnids: A Taxon-vertical Distributional Study of Settling Assemblages From the Panama Basin*. Woods Hole Oceanographic Institution, Woods Hole, USA.

Lipps, J.H., Stoeck, T., Dunthorn, M., 2013. Fossil Tintinnids. In: Dolan, J., Montagnes, D.J.S., Agatha, S., Coats, W., Stoecker, D.K. (Eds.), *The biology and ecology of tintinnid ciliates: models for marine plankton*. Wiley-Blackwell, West Sussex, pp. 186–197.

Lynn, D.H., 2008. *The Ciliated Protozoa: Characterization, Classification, and Guide to the Literature*. 3rd ed. Springer, Dordrecht.

Martín-González, A., Wierzbos, J., Gutiérrez, J.C., Alonso, J., Ascaso, C., 2008. Morphological stasis of protists in lower Cretaceous amber. *Protist* 159, 251–257.

Poinar, G.O., Waggoner, B.M., Bauer, U.-C., 1993. Terrestrial soft-bodied protists and other microorganisms in Triassic amber. *Science* 259, 222–224.

Porter, S.M., Meisterfeld, R., Knoll, A.H., 2003. Vase-shaped microfossils from the Neoproterozoic Chuar Group, Grand Canyon: a classification guided by modern testate amoebae. *J. Paleontol.* 77, 409–429.

Remane, J., 1985. Calpionellids. In: Bolli, H.M., Saunders, J.B., Persh-Nielsen, K. (Eds.), *Plankton stratigraphy. planktic foraminifera, calcareous nannofossils and calpionellids vol. 1*. Cambridge University Press, Cambridge, pp. 555–572.

Rüst, D., 1885. Beiträge zur Kenntniss der fossilen Radiolarien aus Gesteinen des Jura. *Palaeontographica* 36 (N.F. 11), 269–322 + plates 226–245.

Schmidt, A.R., Ragazzi, E., Coppellotti, O., Roghi, G., 2006. A microworld in Triassic amber. *Nature* 444, 835.

- Schönborn, W., Dörfelt, H., Foissner, W., Krientiz, L., Schäfer, U., 1999. A fossilized microcenosis in Triassic amber. *J. Eukaryot. Microbiol.* 46, 571–584.
- Stemans, P., Breuer, P., de Ville de Goyet, F., Marshall, C., Gerrienne, P., 2014. A Givetian tintinnid-like palynomorph from Libya. *Rev. Palaeobot. Palynol.* 203, 3–8.
- Summons, R.E., Walter, M.R., 1990. Molecular fossils and microfossils of prokaryotes and protists from Proterozoic sediments. *Am. J. Sci.* 290-A, 212–244.
- Suzuki, T., Taniguchi, A., 1995. Sinking rate of loricae of some common tintinnid ciliates. *Fish. Oceanogr.* 4, 257–263.
- Takishita, K., Chikaraishi, Y., Leger, M.M., Kim, E., Yabuki, A., Ohkouchi, N., Roger, A.J., 2012. Lateral transfer of tetrahymanol-synthesizing genes has allowed multiple diverse eukaryote lineages to independently adapt to environments without oxygen. *Biol. Direct* 7, 5.
- Tappan, H., 1993. Tintinnids. In: Lipps, J.H. (Ed.), *Fossil prokaryotes and protists*. Blackwell Scientific Publications, Boston, pp. 285–303.
- Tappan, H., Loeblich, A.R.J., 1968. Lorica composition of modern and fossil Tintinnida (ciliate Protozoa), systematics, geological distribution, and some new tertiary taxa. *J. Paleontol.* 42, 1378–1394.